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APPLICATION NO.	FILING DATE	FIRST NAMED INVENTOR	ATTORNEY DOCKET NO.	CONFIRMATION NO.
10/578,860	06/30/2006	Ariel G. Notcovich	227396U	3336
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THE NATH LAW GROUP 112 South West Street Alexandria, VA 22314				
EXAMINER				
LAM, ANN Y				
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Please find below and/or attached an Office communication concerning this application or proceeding.

The time period for reply, if any, is set in the attached communication.

Office Action Summary

Application No.

10/578,860

Applicant(s)

NOTCOVICH ET AL.

Examiner

ANN Y. LAM

Art Unit

1641

Period for Reply -- The MAILING DATE of this communication appears on the cover sheet with the correspondence address --

A SHORTENED STATUTORY PERIOD FOR REPLY IS SET TO EXPIRE 3 MONTH(S) OR THIRTY (30) DAYS, WHICHEVER IS LONGER, FROM THE MAILING DATE OF THIS COMMUNICATION.

- Extensions of time may be available under the provisions of 37 CFR 1.136(a). In no event, however, may a reply be timely filed after SIX (6) MONTHS from the mailing date of this communication.
- If NO period for reply is specified above, the maximum statutory period will apply and will expire SIX (6) MONTHS from the mailing date of this communication.
- Failure to reply within the set or extended period for reply will, by statute, cause the application to become ABANDONED (35 U.S.C. § 133). Any reply received by the Office later than three months after the mailing date of this communication, even if timely filed, may reduce any earned patent term adjustment. See 37 CFR 1.704(b).

Status

- 1) ☒ Responsive to communication(s) filed on 21 August 2009.
- 2a) ☒ This action is **FINAL**. 2b) ☐ This action is non-final.
- 3) ☐ Since this application is in condition for allowance except for formal matters, prosecution as to the merits is closed in accordance with the practice under *Ex parte Quayle*, 1935 C.D. 11, 453 O.G. 213.

Disposition of Claims

- 4) ☒ Claim(s) 29-37, 39 and 41-46 is/are pending in the application.
- 4a) Of the above claim(s) _____ is/are withdrawn from consideration.
- 5) ☐ Claim(s) _____ is/are allowed.
- 6) ☒ Claim(s) 29-37, 39 and 41-46 is/are rejected.
- 7) ☐ Claim(s) _____ is/are objected to.
- 8) ☐ Claim(s) _____ are subject to restriction and/or election requirement.

Application Papers

- 9) ☒ The specification is objected to by the Examiner.
- 10) ☒ The drawing(s) filed on 11 May 2006 is/are: a) ☒ accepted or b) ☐ objected to by the Examiner.
- Applicant may not request that any objection to the drawing(s) be held in abeyance. See 37 CFR 1.85(a).
- Replacement drawing sheet(s) including the correction is required if the drawing(s) is objected to. See 37 CFR 1.121(d).
- 11) ☐ The oath or declaration is objected to by the Examiner. Note the attached Office Action or form PTO-152.

Priority under 35 U.S.C. § 119

- 12) ☐ Acknowledgment is made of a claim for foreign priority under 35 U.S.C. § 119(a)-(d) or (f).
- a) ☐ All b) ☐ Some * c) ☐ None of:
1. ☐ Certified copies of the priority documents have been received.
 2. ☐ Certified copies of the priority documents have been received in Application No. _____.
 3. ☐ Copies of the certified copies of the priority documents have been received in this National Stage application from the International Bureau (PCT Rule 17.2(a)).

* See the attached detailed Office action for a list of the certified copies not received.

Attachment(s)

- 1) ☐ Notice of References Cited (PTO-892)
- 2) ☐ Notice of Draftperson's Patent Drawing Review (PTO-948)
- 3) ☐ Information Disclosure Statement(s) (PTO/SB/08)
Paper No(s)/Mail Date _____
- 4) ☐ Interview Summary (PTO-413)
Paper No(s)/Mail Date _____
- 5) ☐ Notice of Informal Patent Application
- 6) ☐ Other: _____

DETAILED ACTION

Specification

Applicant's amendment to the specification filed August 21, 2009 is acknowledged and has been entered.

The following disclosure is objected to because of informalities: there is no brief description of figures 1A, 1B, 3A-3D, 4A-4D, 5A-5F, 6A-6E, 7A-7F. (It is noted that a brief description of "figure 1" is insufficient to describe figure 1A or 1B.)

Appropriate correction is required.

Claim Objections

Claims 29 and 37 are objected to because of the following informalities.

In claim 29, line 3, "member" should be --members--, since there needs to be more than one member for there to be simultaneous adsorbing. Likewise with line 14, "member" should be --members--.

For the same reason, in claim 37, lines 8 and 12, respectively, "member" should be --members--.

Appropriate correction is required.

Claim Rejections - 35 USC § 112

The following is a quotation of the second paragraph of 35 U.S.C. 112:

The specification shall conclude with one or more claims particularly pointing out and distinctly claiming the subject matter which the applicant regards as his invention.

Claims 29-36 and 43-45 are rejected under 35 U.S.C. 112, second paragraph, as being indefinite for failing to particularly point out and distinctly claim the subject matter which applicant regards as the invention.

Claim 29, lines 22-23, recites "each of the microspots located at a surface between at least two or more the microspots". It is not clear as to which microspots are between which microspots. For examination purposes, the claim will be interpreted to mean that the microspots from which reference data is obtained are between at least two or more microspots that are not used for reference data.

(Claims 30-36 and 43-45 are rejected because they depend from claim 29.)

Claim Rejections - 35 USC § 103

The following is a quotation of 35 U.S.C. 103(a) which forms the basis for all obviousness rejections set forth in this Office action:

(a) A patent may not be obtained though the invention is not identically disclosed or described as set forth in section 102 of this title, if the differences between the subject matter sought to be patented and the prior art are such that the subject matter as a whole would have been obvious at the time the invention was made to a person having ordinary skill in the art to which said subject matter pertains. Patentability shall not be negatived by the manner in which the invention was made.

Claims 29, 30, 33, 36, 37, 41, 42 and 46 are rejected under 35 U.S.C. 103(a) as being unpatentable over Malmqvist et al., 6,200,814, in view of Newgard et al., 6,110,707, and further in view of Lambert, 20060210984.

Malmqvist et al. teach controlling fluid flow over a sensing surface within a flow cell to position the fluid flow over one or more discrete sensing areas within the flow cell (col. 3, lines 14-18.) The term "sensitize" is referred to any process or activation of the sensing area that results in the sensing area being capable of specifically interacting with a desired analyte. The resulting surface is referred to as a "sensitized" area (col. 13, lines 31-35.) For example, the sensing area of the flow cell may be sensitized by immobilization of an analyte-specific ligand (such as antigen, antibody, enzyme, DNA etc. (col. 13, lines 39-58.) Malmqvist et al. disclose that reagents used for sensitizing the sensing area (i.e., immobilization of an analyte-specific ligand) are flowed through different inlets (some parallel, some orthogonal to each other) provided in the flow cell and that different regions can be sensitized with same or different ligands (col. 14, line 20 – col. 15, line 7; and col. 15, line 37 – col. 16, line 27; see also figures 13A-13E.) A gradient of amount of ligands may be provided (col. 14, lines 26-37.) Malmqvist et al. disclose laminar flow techniques to direct the fluid flow but that other techniques may be employed (col. 18, lines 42-47.) Malmqvist et al. disclose that the apparatus can be used to study how multiple biomolecular complexes are formed and how they function, and measuring interactions for the formation of complexes can be by techniques such as SPR (surface plasmon resonance) detection, or fluorescence detection (col. 16, lines 50-59.) Malmqvist et al. disclose use of flows cells for kinetic measurements in general (col. 2, lines 24-31) and it is understood that kinetic measurements are also applicable to the Malmqvist et al. flow cell (see for example col. 8, lines 38-42.)

Thus, as to claims 29, 33, 37, 38, 40, Malmqvist et al. teach, as is also claimed by Applicant, the steps of adsorbing a first binding member at microspots (the different sensitized regions), presenting a second binding member (the binding partner to the immobilized ligand), with a plurality of concentration of both binding members among the plurality of spots (i.e., the gradient of ligands), obtaining data indicative of a binding reaction between the first and second binding members at the spots (i.e., the detection methods such as SPR and fluorescence detection), and it is understood that the method involves kinetic measurements as discussed above. It is noted that the sensitizing step (i.e., step of immobilizing the first ligand binding member) is performed by introducing a flow of the ligand through the flow cell as discussed above. Because the channel of the flow cell must have been formed at some point, and the channels are used to introduce the first binding member for adsorption, there is inherently a step of forming a first channel around a region containing the microspot. While Applicant's specification discloses a flow cell that is mountable on a surface, such limitations are not read into the claims as they are given their broadest reasonable interpretation. Thus, Malmqvist et al.'s activation step (i.e., sensitizing step) meets the limitations of Applicant's step (d) (A) (i) and (ii). It is understood that in the Malmqvist et al. disclosure, excess activating solution (first ligand binding solution) is removed which thus renders the flow cell suitable for subsequent assays.

However, Malmqvist et al. do not teach deactivating the microspot.

Newgard et al. however disclose that in coating a plate with either antigen or antibody, one will generally incubate the wells of the plate with a solution of the antigen

or antibody, either overnight or for a specified period of hours. The wells of the plate will then be washed to remove incompletely adsorbed material. Any remaining available surfaces of the wells are then "coated" with a nonspecific protein that is antigenically neutral with regard to the test antisera. These include bovine serum albumin (BSA), casein and solutions of milk powder. The coating of nonspecific adsorption sites on the immobilizing surface reduces the background caused by nonspecific binding of antisera to the surface (col. 72, lines 4-14.)

Given the teachings of Newgard et al. regarding coating a surface with nonspecific protein to reduce background caused by nonspecific binding, the skilled artisan would utilize the same method taught by Newgard et al., while in regards to wells of a plate, to the invention of Malmqvist et al. to similarly improve the Malmqvist et al. invention to reduce background noise in order to obtain more accurate assay results. Such coating of nonspecific adsorption sites is equivalent to a deactivating step.

Applicant claims *simultaneously* absorbing the first binding member at the discrete spots and also *simultaneously* presenting the second binding member to the first binding member at the discrete spots. Malmqvist et al. disclose that the ligand immobilization may be accomplished by directing fluid containing the ligand over the sensing surface (col. 8, lines 41-47.) A sample flow containing an analyte may then be directed into the flow cell as represented by arrow 1430, such that the sample flow contacts sensing areas 1440, 1450 and 1460. (col. 18, lines 50-53). Since Applicant does not disclose the immobilization of the ligand and analyte such that they are

different from that of simply flowing fluid to contact the discrete spots, the disclosure of Malmqvist et al. in flowing the binding members as described above meet the claimed limitations.

Applicant also claims simultaneously obtaining one or more kinetic parameters indicative of a binding reaction between the first binding member and the second binding member at each of the plurality of microspots to produce a kinetic analysis of the binding, and simultaneously obtaining reference data from a plurality of interspots, each of the microspots located at a surface between at least two or more microspots (interpreted to mean that the reference data spots are between spots with the first and second binding member).

This is disclosed by Lambert in disclosing a microassay chip functionalized with at least one analyte reaction spot, and at least one, and preferably at least two homologous calibration reaction spots arranged in a line (column) perpendicular to the flow of reagent across the surface of the chip, with said at least one analyte reaction spot being arranged in a line (row) with at least one of the calibration reaction spots such that the analyte reaction spot and the calibration reaction spot are parallel with the flow of reagent across the surface of the chip. In a more preferred embodiment, the microassay chip will include a plurality of calibration reaction spots arranged in a series of at least one and preferably at least two or more columns, each column comprised of a homologous population of calibration reaction spots, each calibration reaction spot comprised of preferably peptide nucleic acids, and each of said columns being comprised of spots of a different population of nucleic acid molecules, preferably

peptide nucleic acids. See paragraph 0029. The invention is useful in proteomics for simultaneous analysis of thousands of biomolecular interactions on the surface of the microchip inserted in a flow cell cartridge and provides for normalizing or calibrating for variations in a signal intensity of binding reactions due to variations in reagent flow rate over the surface of the chip that occur as a result of the contact between the flow stream and the surfaces of the flow cell cartridge (paragraph 0002). Various techniques may be used to detect the binding interaction, including surface plasmon resonance (SPR), (paragraph 0004 and 0032.)

It would have been obvious to one of ordinary skills in the art at the time the invention was made to modify the invention of Malmqvist et al. to provide calibration spots in a line between lines of analyte reaction spots because it provides for the advantage of normalizing or calibrating for variations in a signal intensity of binding reactions as taught by Lambert. Providing such a pattern with alternating multiple lines of analyte reaction spots and calibration spots meets the claimed limitation of a plurality of interspots located at a surface between at least two or more microspots. Simultaneous analysis is also disclosed by Lambert (paragraph 002), and the skilled artisan would have recognized the benefits of convenience and efficiency of performing simultaneous reactions. The skilled artisan would have had reasonable expectation of success because both Malmqvist et al. and Lambert disclose patterns of reaction spots in a flow cell and use of surface plasmon resonance as one of the types of detection techniques that can be used.

Applicant also claims that the plurality of bindings carried out do not necessitate a regeneration step. It is disclosed in Applicant's specification in paragraph 0006 that as is known in the art and in commercially available devices, a standard kinetic binding interaction measurement includes washing and regeneration of the probe. That is, in standard kinetic binding interaction measurements, the second binding member (target) is removed so that another concentration of the target is contacted with the probe. Neither Malmqvist et al., nor Newgard et al. nor Lambert disclose that a regeneration step is required, nor is it implied that it is required in order to carry out the disclosed methods, and thus the method discussed above as taught by these references do not necessitate a regeneration step.

As to claim 30, SPR detection is disclosed by Malmqvist et al. (col. 16, lines 50-59.)

As to claim 36, obtaining reference data from a region of the surface not included in a microspot (i.e., another microspot used for control purposes) is disclosed by Malmqvist et al. (col. 14, lines 26-28.)

As to claim 41, forming a second channel perpendicular to the first channel is disclosed (see for example figure 11A, and see such perpendicular flows produced in figures 13A-13E.)

As to claims 42 and 46, a probe array is produced, as shown in figure 13E for example.

Claims 31, 35 and 43 are rejected under 35 U.S.C. 103(a) as being unpatentable over Malmqvist et al., 6,200,814, in view of Newgard et al., 6,110,707, and Lambert, 20060210984, as applied to claim 29 above, and further in view of Lennox et al., 6,478,839.

Malmqvist et al. disclose that the detection method may be SPR (surface Plasmon resonance) but does not specifically disclose the specific type of SPR method claimed, namely that the data indicative of a binding reaction is specifically SPR resonance angle. Lennox et al. disclose this type of SPR technique and also claims it in claim 7, reciting that regarding the SPR detection, the detector includes means for exciting surface plasmons at a plasmon resonance angle that is dependent on the optical properties of the metal film and attached monolayer, and a detector for detecting the shift in plasmon resonance angle produced by binding of ligand-binding agent to said ligand. Because Malmqvist et al. only disclose in general the use of SPR detection for detection of binding, the skilled artisan would look to the art, such as the Lennox et al. patent, for specific types of SPR that would allow for binding detection.

Claims 32, 44 and 45 are rejected under 35 U.S.C. 103(a) as being unpatentable over Malmqvist et al., 6,200,814, in view of Newgard et al., 6,110,707, and Lambert,

20060210984, as applied to claim 29 above, and further in view of Natesan et al., 20020048792.

Malmqvist et al. teach that the flow cell may be used for various assay purposes but do not specifically disclose that the assay is to determine dissociation constant. Natesan et al. however teach in paragraph 0113 that a number of well-characterized assays are available for determining binding affinity, usually expressed as dissociation constant for DNA-binding proteins and the cognate DNA sequences to which they bind. While Malmqvist et al. disclose only in general the use of the flow cell for assay purposes, the skilled artisan would look to the art, such as the Natesan et al. patent, for specific types of assays to be performed.

Claims 34 and 39 are rejected under 35 U.S.C. 103(a) as being unpatentable over Malmqvist et al., 6,200,814, in view of Newgard et al., 6,110,707, and Lambert, 20060210984, and further in view of Siddigi et al., 5,541,113.

The combination of the teachings of Malmqvist et al. and Newgard et al. have been discussed above. However, neither Malmqvist et al. nor Newgard et al. teach activating a microspot by producing an electric field over the microspot.

Siddigi et al. however disclose that it is known that an electric field induces certain chemical reactions (col. 1, lines 51-56.) While the disclosure refers to a chemical reaction that can be detected, rather than for immobilizing a probe, the skilled artisan would recognize that an electric field would induce similar reactions in certain

ligands that may be of interest in order to cause a reaction for immobilization purposes, and thus use of an electric field to induce reactions in the Malmqvist et al. invention would have been obvious to the skilled artisan.

Response

It is noted that Applicant's response filed August 21, 2009 does not include an amendment to the claims nor arguments to the grounds for rejection. The grounds for rejection remains and is made final.

Conclusion

THIS ACTION IS MADE FINAL. Applicant is reminded of the extension of time policy as set forth in 37 CFR 1.136(a).

A shortened statutory period for reply to this final action is set to expire THREE MONTHS from the mailing date of this action. In the event a first reply is filed within TWO MONTHS of the mailing date of this final action and the advisory action is not mailed until after the end of the THREE-MONTH shortened statutory period, then the shortened statutory period will expire on the date the advisory action is mailed, and any extension fee pursuant to 37 CFR 1.136(a) will be calculated from the mailing date of the advisory action. In no event, however, will the statutory period for reply expire later than SIX MONTHS from the mailing date of this final action.

Any inquiry concerning this communication or earlier communications from the examiner should be directed to ANN Y. LAM whose telephone number is (571)272-0822. The examiner can normally be reached on Mon.-Thurs. 9-7:30.

If attempts to reach the examiner by telephone are unsuccessful, the examiner's supervisor, Mark Shibuya can be reached on 571-272-0806. The fax phone number for the organization where this application or proceeding is assigned is 571-273-8300.

Information regarding the status of an application may be obtained from the Patent Application Information Retrieval (PAIR) system. Status information for published applications may be obtained from either Private PAIR or Public PAIR. Status information for unpublished applications is available through Private PAIR only. For more information about the PAIR system, see <http://pair-direct.uspto.gov>. Should you have questions on access to the Private PAIR system, contact the Electronic Business Center (EBC) at 866-217-9197 (toll-free). If you would like assistance from a USPTO Customer Service Representative or access to the automated information system, call 800-786-9199 (IN USA OR CANADA) or 571-272-1000.

/Ann Y. Lam/
Primary Examiner, Art Unit 1641